

PHARMACOLOGY AND TOXICOLOGY

Bioactive 5-Reduced 16 α ,17 α -Cyclohexanoprogesterone Derivatives Weakly Interact with Proteins of the Soluble Uterine Fraction

A. N. Smirnov, E. V. Pokrovskaya, I. S. Levina*, L. E. Kulikova*,
A. V. Kamernitskii*, and V. P. Shevchenko**

Translated from *Byulleten' Eksperimental'noi Biologii i Meditsiny*, Vol. 131, No. 3, pp. 293-296, March, 2001
Original article submitted October 2, 2000

We studied competitive activities of 16 α ,17 α -cyclohexano-5 α - and 5 β -dihydroprogesterone in replacing ^3H -progesterone and ^3H -16 α ,17 α -cyclohexano-6 α -methylprogesterone from protein complexes. Direct binding of ^3H -5-reduced derivatives with proteins of soluble fractions from rat and rabbit uteri was also assayed. C_d values for 5-reduced derivatives were in the micro- or submicromolar range. The data suggest that biological effects of these analogues are not mediated via soluble uterine receptors.

Key Words: *progestins; steroids; analogues; receptors; binding*

Pregna-D'-pentaranes are progesterone derivatives containing additional 16 α ,17 α -carbocycles and possessing high progestogen activity (in some cases, selective) [1,4,5]. Their affinity for progesterone receptors is comparable with that of progesterone. 5-Reduced pregna-D'-pentarane derivatives are analogues of 5 α /5 β -progesterone metabolites. Progestogen activity of 5-reduced progesterone metabolites is much lower than that of the initial hormone: 5 α -dihydroprogesterone derivatives only partially retain, while 5 β -dihydroprogesterone metabolites completely lose their hormonal activity, which correlates with the affinity of these compounds for progesterone receptors [3]. These data suggest that 5 α /5 β -reduction of progesterone analogues having additional D' cycloalkane rings would produce similar changes. The Clauberg—McFeil test showed that 16 α ,17 α -cyclohexano-5 α /5 β -dihydroprogesterones possess no progestogen activity [1]. Both

compounds are potent antigestogens inhibiting progesterone-induced stimulation of endometrial cell proliferation. 5 α -Reduced analogues counteract the pregnancy-maintaining effect of progesterone. It should be emphasized that the effective dose of 5 α /5 β -reduced pregna-D'-pentaranes is comparable with that of the natural hormone [8]. We hypothesize, that either reduction of A rings in pregna-D'-pentaranes does not decrease their affinity for progesterone receptors or these compounds act by a nonclassical pathway. The protein isolated from soluble fractions of rat uterus and specifically binding pregna-D'-pentaranes is probably involved in this tentative pathway [10]. Probably, there are some specific receptors for 5-reduced pregna-D'-pentaranes.

The interaction of 16 α ,17 α -cyclohexano-5 α /5 β -dihydroprogesterones with soluble uterine fraction proteins was studied by competitive assay and direct binding of ^3H -labeled compounds.

MATERIALS AND METHODS

We studied 1,2,6,7- ^3H -progesterone (specific radioactivity 86 Ci/mmol, St. Petersburg), 6 α -methyl-1,2- ^3H -

Laboratory of Endocrinology, Department of Biology, M. V. Lomonosov Moscow State University; *Group of Steroids and Oxylipins, N. D. Zelinskii Institute of Organic Chemistry, Russian Academy of Sciences; **Laboratory of Isotopically Modified and Physiologically Active Substances, Institute of Molecular Genetics, Russian Academy of Sciences

16 α ,17 α -cyclohexanoprogesterone (43 Ci/mmol), 1,2,3',4'- 3 H-16 α ,17 α -cyclohexano-5 α -dihydroprogesterone (83 Ci/mmol), and 1,2,3',4'- 3 H-16 α ,17 α -cyclohexano-5 β -dihydroprogesterone (83 Ci/mmol) [9]. Unlabeled compounds were obtained as described previously [8]. Other reagents were from Sigma.

Experiments were performed on adult female rats weighing 180-200 g. The animals were intramuscularly injected with 1 μ g estradiol in 0.2 ml propylene glycol for 4 days. The rats were decapitated 1 day after the last injection. The uteri were removed and homogenized in 10 mM Tris-HCl buffer (pH 7.5) containing 10 mM KCl, 1 mM EDTA, 1 mM dithiothreitol, 0.5 mM phenylmethylsulfonyl fluoride, and 30% glycerol. All procedures were performed at 0-4°C. The supernatant (cytosolic fraction) containing 5-8 mg/ml protein was obtained by centrifugation of the homogenate at 50,000g for 1 h. Aliquots of the cytosolic fraction (100 μ l) were incubated with 3-4 mBq 3 H-progesterone (in the presence of 3 μ M hydrocortisone) and unlabeled competitors in concentrations of 0-10 μ M for 20 h to study the interaction of test compounds with progesterone receptors. 3 H-6 α -Methyl-16 α ,17 α -cyclohexanoprogesterone (6 mBq) was used as a labeled ligand to study the interaction of steroids with the protein specifically binding pregna-D'-pentanenes. The reaction was performed in the presence of 127 nM progesterone inhibiting binding with progesterone receptors and varying concentrations of competitors. Direct interaction of 3 H-16 α ,17 α -cyclohexano-5 α / β -dihydroprogesterones (4-6 mBq) with proteins was studied in the presence of unlabeled steroids in vari-

ous concentrations. Protein-bound and free ligands were separated with dextran-coated activated charcoal for 5 min. After centrifugation, radioactivity in supernatant aliquots was measured with an efficiency of 20%. This method and calculations of the equilibrium dissociation constant (C_d) and relative competitive activity (RCA) were described elsewhere [4,5]. Protein content was measured by Coomassie staining [7].

RESULTS

5-Reduced progesterone analogues weakly interact with progesterone receptors and pregna-D'-pentanene-binding protein (Table 1, Fig. 1). C_d for these compounds surpassed 1 μ M, which was 2.5 and 1.5 orders of magnitude higher than that for the corresponding high-affinity protein ligands. It should be emphasized that 5 β -reduced derivative had greater C_d in relation to 2 proteins compared to that of 5 α -reduced analogue, which indicated similar structure of steroid-binding sites in both proteins. Direct binding of 3 H-labeled 16 α ,17 α -cyclohexano-5 α / β -dihydroprogesterones by soluble uterine fraction proteins was characterized by C_d 0.2 and 2.3 μ M for 5 α - and 5 α / β -analogues, respectively. These results are in contradiction with the assumption that rat uterus contains specific high-affinity soluble receptors for 5 α / β -analogues. Experiments on rabbits demonstrated high antiprogesterone activity of 16 α ,17 α -cyclohexano-5 α / β -dihydroprogesterones [8]. It cannot be excluded that gestogen receptors in rabbits differ from those in rats. In special experimental series we studied soluble uterine

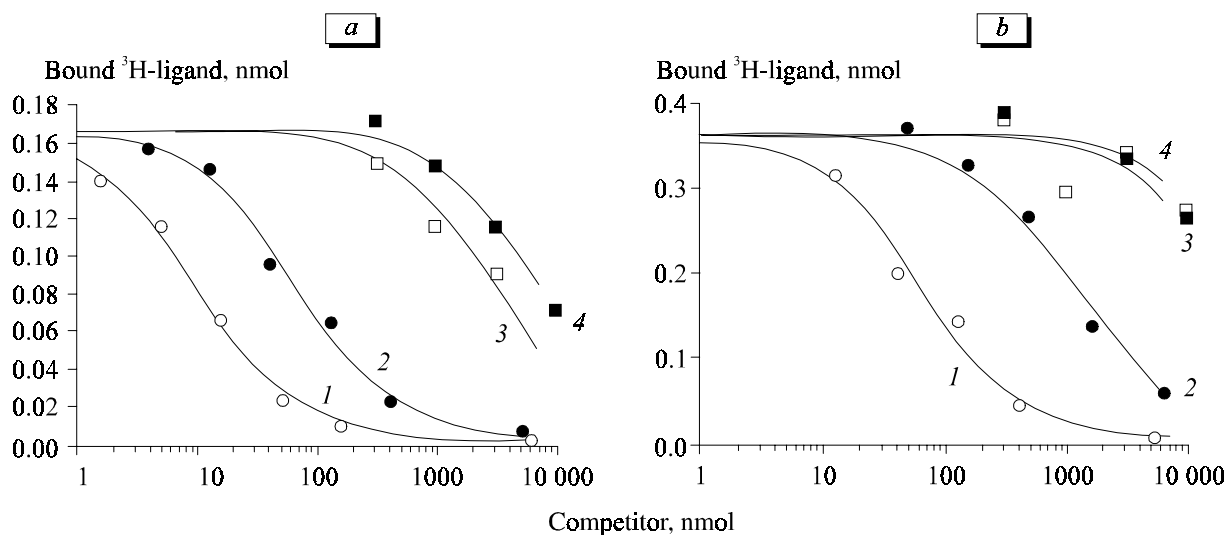


Fig. 1. Competitive assay of interaction between 5-reduced 16 α ,17 α -cyclohexanoprogesterone analogues and progesterone receptor (a) or protein from soluble rat uterine fraction specifically binding pregna-D'-pentanenes (b). a) progesterone (1), 16 α ,17 α -cyclohexano-6 α -methylprogesterone (2), 16 α ,17 α -cyclohexano-5 α -dihydroprogesterone (3), and 16 α ,17 α -cyclohexano-5 β -dihydroprogesterone (4). b) 16 α ,17 α -cyclohexano-6 α -methylprogesterone (1), progesterone (2), 16 α ,17 α -cyclohexano-5 α -dihydroprogesterone (3), and 16 α ,17 α -cyclohexano-5 β -dihydroprogesterone (4).

TABLE 1. Interaction of 5-Reduced 16 α ,17 α -Cyclohexanoprogesterone Analogues with Progesterone Receptor (I) and Protein from Soluble Rat Uterine Fraction (II) Specifically Binding Pregna-D'-Pentaranes

³ H-ligand/protein	Unlabeled competitor	C _d , nmol	RCA
Progesterone/I	Progesterone	4.8±1.1 (3)	1
	16 α ,17 α -Cyclohexano-6 α -methylprogesterone	36.0 (2)	0.167
	16 α ,17 α -Cyclohexano-5 α -methylprogesterone	1790±460 (3)	0.0030±0.0009
	16 α ,17 α -Cyclohexano-5 α -dihydroprogesterone	7380±3940 (3)	0.0012±0.0006
16 α ,17 α -Cyclohexano-6 α -methylprogesterone/II	16 α ,17 α -Cyclohexano-6 α -methylprogesterone	56.9±31.9 (3)	1
	Progesterone	1100±595 (3)	0.046±0.008
	16 α ,17 α -Cyclohexano-5 α -dihydroprogesterone	7940±5130 (3)	0.027±0.014*
	16 α ,17 α -Cyclohexano-5 β -dihydroprogesterone	345000±327000 (3)	0.0014±0.0009

Note. Number of measurements is shown in parentheses. * $p < 0.05$ compared to 5 β -analogue.

fraction from estrogenized rabbits. 5-Reduced derivatives had low affinity for progesterone receptors: C_d for 16 α ,17 α -cyclohexano-5 α - and 5 β -dihydroprogesterone were 1 and 0.68 μ mol, respectively; RCA (compared to progesterone) were 0.0179 and 0.0226, respectively. Two experiments with rabbit samples and ³H-16 α ,17 α -cyclohexano-5 β -dihydroprogesterone as a labeled ligand showed that C_d and RCA were 590 nmol and 0.06, respectively. Thus, 16 α ,17 α -cyclohexano-5 α / β -dihydroprogesterones probably do not have high-affinity soluble receptors in the uterus of estrogenized rats and rabbits. High biological activity of these compounds probably results from their low clearance due to low-intensity metabolism and/or strong binding to blood transport proteins. It can not be excluded that the effects of these substances are realized via extragenomic membrane receptors, whose hormonal specificity differs from that of the classic soluble gestogen receptors [6]. It should be emphasized that RCA of 5-reduced pregna-D'-pentaranes in relation to progesterone receptors in the uterus of pregnant rats [2] far surpassed the values obtained in our experiments. This contradiction is probably related to peculi-

arities of physiological regulation of receptor properties, e.g., their phosphorylation.

This work was supported by the Russian Foundation for Basic Research (grant No. 99-03-33033).

REFERENCES

1. A. V. Kamernitzky and I. S. Levina, *Khim.-Farm. Zh.*, **25**, No. 10, 4-16 (1991).
2. E. N. Kareva, A. V. Kamernitzky, I. S. Levina, et al., *Eksp. Klin. Farmakol.*, **62**, No. 5, 25-27 (1999).
3. V. B. Rozen and A. N. Smirnov, *Receptors and Steroid Hormones* [in Russian], Moscow (1981).
4. A. N. Smirnov, E. V. Pokrovskaya, V. P. Shevchenko, et al., *Biokhimiya*, **61**, No. 9, 1279-1287 (1998).
5. A. N. Smirnov, E. V. Pokrovskaya, V. P. Shevchenko, et al., *Bioorg. Khimiya*, **25**, No. 10, 774-781 (1999).
6. P. F. Blackmore, J. F. Fisher, C. H. Spilman, and J. E. Bleasdale, *Mol. Pharmacol.*, **49**, 727-739 (1996).
7. M. M. Bradford, *Analyt. Biochem.*, **72**, No. 2, 248-254 (1976).
8. I. S. Levina, G. V. Nikitina, L. E. Kulikova, and A. V. Kamernitzky, *Rus. Chem. Bull.*, **44**, No. 3, 547-550 (1995).
9. V. P. Shevchenko, I. Yu. Nagaev, A. V. Potapova, et al., *J. Labelled Cpd. Radiopharm.*, **41**, 919-925 (1998).
10. A. N. Smirnov, E. V. Pokrovskaya, G. S. Kogteva, et al., *Steroids*, **65**, No. 3, 163-170 (2000).